

A SEMINAR PAPER ON
ROLE OF MICROORGANISM IN SOIL FERTILITY AND CARBON
SEQUESTRATION

Course Title: Seminar

Course Code: SSC 598

Summer

SUBMITTED TO:

Course Instructors	Major Professor
1. Dr. Md. Mizanur Rahman Professor BSMRAU	Dr. Md. Mizanur Rahman Professor , Dept. of Soil Science BSMRAU
2. Dr. A. K. M. Aminul Islam Professor BSMRAU	
3. Dr. Md. Rafiqul Islam Professor BSMRAU	
4. Dr. Dinesh Chandra Shaha Assistant Professor BSMRAU	

SUBMITTED BY:

ShahidaArofi

MS Student

Reg. No.: 13-05-2954

Department of Soil Science

BANGABANDHU SHEIKH MUJIBUR RAHMAN AGRICULTURAL UNIVERSITY

SALNA, GAZIPUR 1706

Abstract

Soil microorganism are vital part of soil and microbial mass vary with soil depth. So availability of nutrients also vary with soil depth. Almost every chemical reactions in soil is done by microbes and make nutrients available for plant and crops uptake. They play an active role in soil fertility hence all of the bio-geochemical cycle mediated by microbes such as nitrogen ,phosphorus, carbon cycle etc. By the decomposition of organic matter, microbes recycle the nutrients in soil. Nitrogen fixation is occurred by microbes either free living or non-freeliving microorganism. *Rhizobium* is the most common symbiotic bacteria which can fix nitrogen biologically. Some bacteria such as *Micrococcus* spp., *Enterobacter aerogens*, *Pseudomonas capacia* and some funji like *Aspergillus niger*, *A. flavus*, *A. japonicas*, *Penicillium* sp., and also actinomycetes like *Streptomyces* can solubilize p and make available for plant. Micorrhiza help plant to uptake nutrients from deeper horizon specially p. Micorrhiza mediated nontoxification of some inorganic pollutant like As, Cd, Pb ,Cu. Organic fertilizer(Compost) and biofertilizer are made by microbes helps to improve soil fertility and these are very eco-friendly. In recent past there is an another important fuction of microbes has been invented, that is role of microbes in carbon sequestration. Soil is a large sink for storing atmospheric carbon. Since carbon is the main reason for global warming ,microbes play a great role by storing carbon in soil. Some ectomicorrhiza and bacteria can fix carbon. Carbon can persist in humas for long time by the humification process.

Content

Topics	Page No.
Introduction	01-02
Objectives	02
Materials & Methods	03
Review & Discussion of Findings	04-21
Conclusions	22
References	23-25

List of Tables

Serial no.	Title of Tables	Page no.
1	Essential plant nutrient in soil and their available form	04
2	Distribution of Microorganisms in soil according to soil depth	05
3	Some estimates of the amount of nitrogen fixed on a global scale	06
4	A short list of <i>Rhizobium</i> species that fix nitrogen symbiotically and their corresponding hosts	07
5	Increase in rice grain yield and estimated amounts of fixed N ₂ by different N ₂ fixing systems	07
6	Phosphorus solubilizing microorganisms	08
7	Some amounts of P solubilized by selected species o microorganism	08
8	Total P accumulation in cultures of different bacterial species grown on insoluble mineral phosphate substrates	09
9	Inoculation ct of <i>Bacillus</i> sp. on seed yield and plant height of cotton	10
10	Inoculation effect of <i>Bacillus</i> sp. on NP content in cotton leaves	10
11	Effects of <i>Enterobacteragglomerans</i> (PSRB) and <i>Glomus.etunicatum</i> (AMF) inoculation on tomato plant growth and P uptake (75 days after inoculation)	11
12	Mycorrhiza increase soil fertility	13
13	AR ($\mu\text{g mg}^{-1}$) of macroelements (P, Ca and Mg) and microelements (Cu, Zn and Mn) in mycorrhizal (+M) and nonmycorrhizal (-M) wheat plants	13
14	Mycorrhiza mediated nontoxification of some inorganic pollutants in soils their mechanisms	13
15	Average yields by treatment with compost in kg/ha for 5 crops	15
16	Microorganisms and uptake of heavy metals	16
17	Effect of inoculum size on the lindane removal by <i>Streptomyces</i> sp.in sterile soil samples, after 4 weeks of incubation	16
18	Some commonly used biofertilizers with their active ingredient	17
19	Fertilizer saved when biofertilizer applied and their recommended crops	18

List of Figures

Serial no	Name of the figures	Page no.
1	Clover root bearing naturally occurring nodules of <i>Rhizobium</i>	06
2	Schematic diagram of soil phosphorus mobilization and immobilization by bacteria	19
3	Available P status after 30 days of sowing cotton (inoculated with <i>Bacillus sp</i>)	11
4	Available P status after 30 days of sowing cotton (inoculated with <i>Bacillus sp</i>)	12
5	Response curve was produced by growing mycorrhizal and nonmycorrhizal plants <i>Cassia</i> at a range of soil P levels.	12
6	Tree bearing the orange-yellow coloured nodules (arrowheads) containing <i>Frankias</i>	14
7	Response of a soybean crop in Hawaii to inoculation with rhizobia biofertilizer with various levels of phosphorus fertilizer added	27
8	Plant associated microorganism and carbon	19
9	Fruting body ectomycorrhizal fungi from genus <i>Amantia</i>	20
10	Soil carbon storage by Humification	20

CHAPTER I

INTRODUCTION

Microorganisms play very useful role in soil fertility. Usually people think that microbes are agents of disease, but, they perform many other beneficial functions in soil. The beneficial microorganisms help in the decomposition of organic residues, toxic substances and other pollutants and add to the soil fertility (Afzal, 2012). The role they play in improving the soil fertility has become a subject of more investigations during the recent past.

Capacity of soil to supply the essential plant nutrients in available form and in a proper balance for healthy plant growth is called soil fertility. Soil fertility depends on the presence of inorganic substances, organic substances, water and air, as well as, on the presence of microbes (Basta, 2011).

A renowned microbiologist Jacob Lipman, remarked, “A soil lack of microorganisms is dead soil”. Soil is not a dead mass. Fertile soil contains a wide variety of microbes which include different types of bacteria, fungi, protozoa, algae and actinomycetes. Mostly they are found in rhizosphere where they decompose organic matter to humus which is the store house of nutrients and influences soil fertility. Microorganisms are responsible for making up numerous transformations, which changes plant nutrients to readily available forms and make and stabilize desirable soil structure for luxuriant plant growth. Phosphorus Solubilizing bacteria and fungi play an important role in converting insoluble phosphatic compound such as rock phosphate, bone meal etc particularly the chemically fixed soil phosphorus into available form. Mycorrhiza plays a very important role in supplying nutrients to the crop plants by improving soil fertility through increasing root surface area. Soil organic matter holds more than 95% of soil nitrogen, 5-60% of total phosphorus and 30% of soil sulfur. The benefits of soil microorganisms are realized through soil management practices or cropping systems that maximize their activities in their soil habitat, composting and application of effective microorganisms as inoculants or biofertilizers (Omar, 2009).

Microorganisms have also role in carbon sequestration. Which a most important topic is in now days, since Amount of carbon is increasing day by day. Over the past 150 years amount of carbon has increased by 30%.

We know that there is direct relationship between increased levels of carbon and global warming. So how can we reduce carbon from atmosphere it's a burning question. One of the granted proposals is carbon storage in soil which is one part of carbon sequestration.

Carbon sequestration refers the process of removal of carbon the atmosphere and storage in soil, ocean and vegetation. Although oceans store most of the Earth's carbon, soils contain approximately 75% of the carbon pool on land. Carbon cycle is dominated by the balance between photosynthesis and respiration. Carbon is transferred from the atmosphere to soil by 'carbon-fixing' autotrophic organisms; they also make carbon dioxide (CO₂) into organic material. The objectives of the study are-

- To review the role that play microorganisms in N-fixation and P-solubilization,
- To review the role of Mycorrhiza in improving soil fertility,
- To review the role of microorganisms in heavy metal uptake and pollutants degradation and
- To review role of microorganism in soil carbon sequestration

CHAPTER II

MATERIALSAND METHODS

This seminar paper is exclusively a review paper so all of the information has been collected from the secondary sources. During preparation of this paper I went through various relevant books, journals, proceedings, reports, publications etc. Findings related to my topic have been reviewed with the help of the library facilities of Bangabandhu Sheikh MujiburRahman Agricultural University (BSMRAU) and Bangladesh Agricultural Research Institute (BARI). Information also collected from Bangladesh Agricultural Research Council (BARC). I have also searched related internet web sites to collect information. I got valuable suggestion and information from my major professor and course instructors. After collecting all the available information, I myself compiled and prepared this seminar paper.

CHAPTER III

REVIEW AND DISCUSSION OF THE FINDINGS

The soil fertility improving functions of soil microorganisms include release of plant nutrients such as P,K and Zn from insoluble inorganic forms, decomposition of organic residues and release of nutrients, formation of beneficial soil humus by decomposing organic residues and through synthesis of new compounds ,production of plant growth promoting compounds and improvement of plant nutrition through symbiosis.

Fertile soil contains all of essential nutrients require for plant growth and development in available form. The conversion of complex molecular compounds into ionic forms is carried out by microorganisms either directly or indirectly. (Table1).

Table1.Essential plant nutrient in soil and their available form

Element name	Chemical symbol	Available form to plants	%Concentration in dry tissue
Nitrogen	N	NO_3^- , NH_4^{4+}	1.5
Potassium	K	K^+	1.0
Phosphorus	P	H_2PO_4^- , HPO_4^{2-}	0.2
Calcium	Ca	Ca^{2+}	0.5
Magnesium	Mg	Mg^{2+}	0.2
Sulfur	S	SO_4^{2-}	0.1
Zinc	Zn	Zn^{2+}	0.002
Iron	Fe	Fe^{2+} , Fe^{3+}	0.01
Manganese	Mn	Mn^{2+}	0.005
Copper	Cu	Cu^{2+}	0.0006
Boron	B	H_3BO_3	0.002
Molybdenum	Mo	MoO_4^{2-}	0.00001
Chlorine	Cl	Cl^-	0.01
Oxygen	O	O_2 , H_2O	45
Carbon	C	CO_2	45
Hydrogen	H	H_2O	6

Source: FAO, 2004.

Distribution of microorganisms in soil

There are many microorganisms live in soil and they perform many activities. Among them the number of bacteria per gram soil is highest, then actinomycetes and fungi. The number of microorganisms varies according to depth of soil (Table2). The highest number of bacteria (7800000+195000) lived in top soil which is much higher (only 1000) than sub soil or deeper soil.

Table2.Distribution of Microorganisms in soil according to soil depth

Depth	Microorganisms per gram of soil			
CM	Aerobic Bacteria	Anaerobic Bacteria	Actinomyces	Fungi
3-8	7800000	1950000	2080000	119000
20-25	1800000	379000	245000	50000
35-40	472000	98000	49000	14000
65-75	10000	1000	5000	3000
135-	100	400		3000

Source: Alexander, 2008

Nitrogen fixation by microorganisms

A relatively small amount (<1% of total) of ammonia is produced by lightning. Some ammonia also is produced industrially (about 50×10^6 MT) by the Haber-Bosch process, using an iron-based catalyst, very high pressures and fairly high temperature ($400-550^{\circ}\text{C}$ and 200 atm). But the major conversion of N_2 into ammonia, and thence into proteins, is achieved by microorganisms in the process called nitrogen fixation. The total biological nitrogen fixation (about 175×10^6 MT) is estimated to be twice as much as the total nitrogen fixation (about 80×10^6 MT) by non-biological processes (Table3).

Table3. Some estimates of the amount of nitrogen fixed on a global scale

Type of fixation	N ₂ fixed (10 ¹² g per year, or 10 ⁶ metric tons per year)
Non-biological	
Industrial	about 50
Combustion	about 20
Lightning	about 10
Total	about 80
Biological	
Agricultural land	about 90
Forest and non-agricultural land	about 50
Sea	about 35
Total	about 175

Source: Kennedy *et al.*, 2009.

The nitrogen-fixing organisms

All the nitrogen-fixing organisms are prokaryotes (bacteria). Some of them live independently of other organisms - the so-called free-living nitrogen-fixing bacteria. Others live in intimate symbiotic associations with plants or with other organisms (e.g. *Rhizobium*).

Symbiotic nitrogen fixation

Legume symbioses

The most familiar examples of nitrogen-fixing symbioses are the root nodules of legumes (peas, beans, clover, etc.).



Fig1. Clover root bearing naturally occurring nodules of *Rhizobium*.

Table4. A short list of *Rhizobium* species that fix nitrogen symbiotically and their corresponding hosts

Rhizobium species	Host plants
<i>Bradyrhizobiumjaponicum</i>	<i>Glycine max</i> (soybean)
<i>R. phaseoli</i>	<i>Phaseolus vulgaris</i> (common bean)
<i>R. meliloti</i>	<i>Medicago sativa</i> (alfalfa)
<i>R. trifolii</i>	<i>Melilotus</i> sp. (sweet clovers)
<i>R. trifolii</i>	<i>Trifolium</i> sp.

Source: Peoples *et al.*, 2007

N₂– Fixators

When *Azolla-Anabaena* inoculated to rice at field conditions showed yield increase 1.5 t/ha which is 50% higher than control field. Similarly *Cyanobacteria*, *Azotobacter*, *R.leguminosarum* inoculated field showed yield increase 29%, 20% and 2-22% (Table5) respectively

Table 5. Increase in rice grain yield and estimated amounts of fixed N₂ by different N₂fixing systems

N ₂ – Fixators	Increase in rice yield		Estimated amount of N ₂
	Amount	(%)	
<i>Azolla-Anabaena</i>	1.5 t ha ⁻¹	50	48.2 kg ha ⁻¹
<i>Cyanobacteria</i>	1.4 t ha ⁻¹	29	24.2 kg ha ⁻¹
<i>Azotobactersp</i>	0.4-0.9 t ha ⁻¹	7-20	11–15 kg ha ⁻¹
<i>Rhizobium leguminosarum</i>	0.6–7.9 g pot ⁻¹	2-22	23–31 mg

Source: Ladaha *et al.*, 2004

Phosphorus-solubilizing microorganisms

Microorganisms solubilize P through production of low molecular weight organic acids in which hydroxyl and carboxyl groups chelate cations that are associated with complexed forms of P (Ca, Al and Fe) thus rendering phosphate soluble in both basic and acid soils and directly dissolve mineral phosphates from Al-P and Fe-P complexes as a result of anion exchange of PO₄³⁻ with acid anion. The organic acids and proton release mechanisms by microorganisms also decrease the pH in basic soils and thus solubilize P from the calcium phosphate (Ca-P).

Substantial amounts of P solubilized by some selected microbial species/strains as reported by a number of authors are presented in Table6.

Table6. Phosphorus solubilizing microorganisms

Bacteria	Fungi	Actinomycetes
<i>Enterobacteraerogenes</i>	<i>Aspergillusflavus</i>	
<i>Pseudomonas cepacia</i>	<i>Penicilliumradicum</i>	
<i>Bacillus licheniformis</i>	<i>A. Niger</i>	<i>Streptomyces</i>
<i>Micrococcus spp.</i>	<i>Penicillium variable</i>	
<i>Enterobacterintermedium</i>	<i>A. Japonicas</i>	

Source: Hayat et al., 2007.

In a study scientist showed that *Arthrobactersp.* solubilized P about 550 mg/L in their life time which is higher than that of *Aspergillusniger*(400 mg/L) ,*Chryseobacteriumsp.*(289.8 mg/L),*Burkholderiasp.*(167.2 mg/L) and*Pantoeasp.*(479 mg/L). Substantial amounts of P solubilized by some selected microbial species/strains as reported by a number of authors are presented in Table 7.

Table7. Some amounts of P solubilized by selected species of microorganism

Mechanism	Amount of P Solubilized (mg/L)	Microbial species/strains	References
Production of organic acids(citric acid and lactic acid)	519.7	<i>Arthrobactersp.</i>	Okokoet al.,2007
Production of organic acids	400	<i>Aspergillusniger</i>	Edwards et al.,2009
Production of organic acids	293	<i>Enterobacter cloacae</i>	Gilleret al.,2012
Production organic acids (citric acid)	289.8	<i>Chryseobacteriumsp.</i>	Sangingaet al.,2012
Production of acids;	167.2	<i>Burkholderiasp.</i>	Oppermanet al.,2008
Production of organic acids	479	<i>Pantoeasp.</i>	Rannaegymanet al.,2009

Phosphorus Mobilization

Microorganisms play critical roles in soil P dynamics including mineralization and immobilization of organic P. The positive influences of soil microorganisms are on mineralization of organic P and solubilization of P from its fixed or precipitated forms including P from rock phosphate.

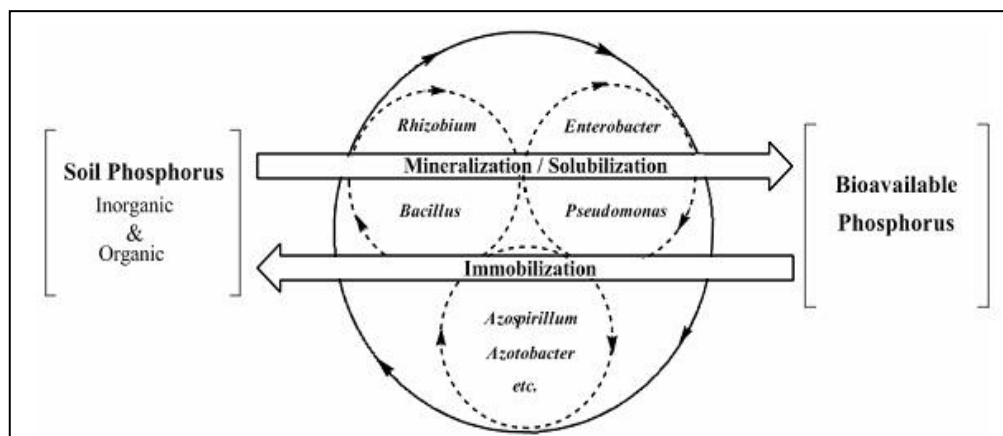


Fig2.Schematic diagram of soil phosphorus mobilization and immobilization by bacteria.**Source: Khan et al., 2012.**

There are many bacteria that amass P from insoluble mineral phosphate substrates. A study showed that some bacteria such as *Bacillus polymyxa*, *Pseudomonas striata*, *Bacillus circulans* etc grow on insoluble substrate (Hydroxyapatite) could amass 87,143 and 65ppm P respectively (Table 8).

Table 8. Total P accumulation in cultures of different bacterial species grown on insoluble mineral phosphate substrates

Bacterial strain	Substrate			Reference
	Ca ₃ (PO ₄) ₂ (ppm)	Hydroxyapatite (ppm)	Rock phosphate(ppm)	
<i>Bacillus polymyxa</i>	116	87	17	Bishop et al., 2009
<i>Pseudomonas striata</i>	156	143	22	Riccio et al., 2008
<i>Burkholderiacepacia</i>	35	Nd	Nd	Suslovet al., 2003
<i>Rhizobium sp.</i>	Nd	300	Nd	Kloeppe et al., 2003
<i>Rhizobium meliloti</i>	Nd	165	Nd	Bishop et al., 2009
<i>Bacillus circulans</i>	11	65	13	Glick et al., 2002

Source: www.bashanfoundation.org

Effect of *Bacillus* sp. on cotton yield and plant height

Phosphate solubilizing *Bacillus* sp enhanced the seed cotton yield and plant height (Table 9). The highest seed cotton yield was produced with *Bacillus* inoculation (1733.3 kg ha⁻¹) at 90 kg P ha⁻¹. Bacterial inoculation produced higher seed cotton yield at all P levels compared to their general control condition. Similarly, bacterial inoculation produced also higher plant height than their usual controls. Data regarding NP content in cotton leaves was put on (Table 10). Inoculation with *Bacillus* sp produced highest N-content (1.707%) at 90 kg P ha⁻¹. Higher N and P-content in cotton leaves was observed with inoculated treatments as compared to un-inoculated ones.

Table9. Inoculation effect of *Bacillus* sp. on seed yield and plant height of cotton

Treatments	Seed cotton Yield (kg ha ⁻¹)		Plant Height (cm)	
kg P ha-1	Un-inoculated	Inoculated	Un-inoculated	Inoculated
30	1377.7	1489.0	152.5	156.5
60	1544.3	1666.7	154.5	158.7
90	1611.3	1733.3	157.2	160.2

Source: Qureshiet al., 2012

Table10. Inoculation effect of *Bacillus* sp. on NP content in cotton leaves

Treatments	N-content (%)		P-content (%)	
kg P ha-1	Un-inoculated	Inoculated	Un-inoculated	Inoculated
30	1.627	1.660	0.227	0.243
60	1.660	1.683	0.250	0.273
90	1.670	1.707	0.257	0.277

Source: Qureshiet al., 2012

Inoculation with *Bacillus* sp enhanced the availability of P at every case than un-inoculated treatments.

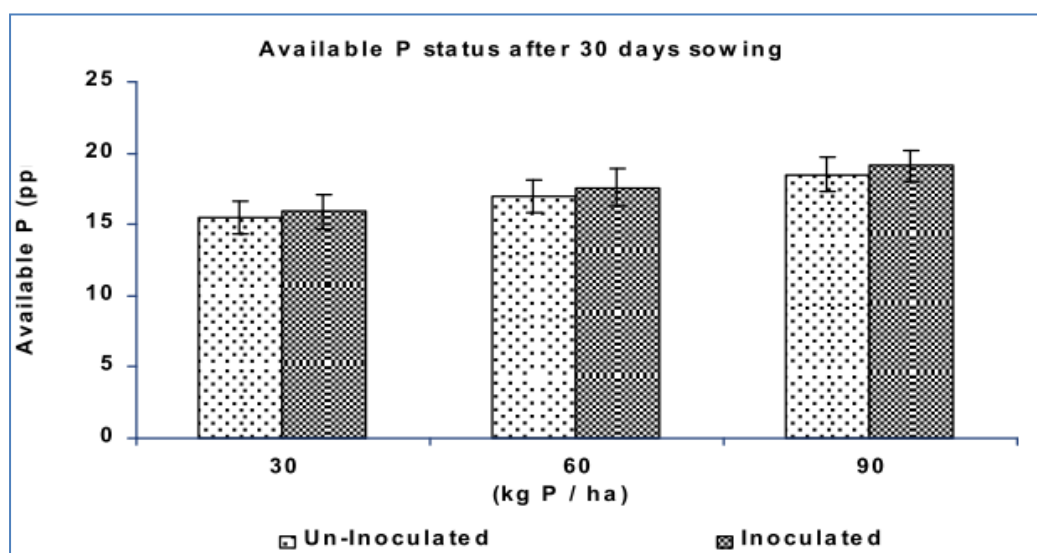


Fig3.

Available P status after 30 days of sowing cotton (inoculated with *Bacillus sp.*).

Effects of PSRB and AMF inoculation on tomato plant

PSRB and AMF significantly enhanced the growth of tomato plant and increase uptake of P (Table 11). PSRB and AMF inoculation produced higher growth and P uptake compared to their general control. Inoculation with PSRB+AMF togetherly produced highest P-content in both shoot (134.41g/plant) and root (16.7g/plant) (Table 11).

Table 11. Effects of *Enterobacter agglomerans* (PSRB) and *Glomus etunicatum* (AMF) inoculation on tomato plant growth and P uptake (75 days after inoculation)

Treatment	Shoot dry weight (g per plant)	Root dry weight (g per plant)	Total P (g per plant)	
			Shoot	Root
Control	4221	429	11.6	2.5
PSRB	4849	510	12.5	3.2
AMF	4762	557	13.7	3.6
PSRB + AMF	54,56	6,77	27.44	16.7

Source: Kim Kim et al, 2009.

Fertility improvement by micorrhizal fungi

Mycorrhizas are refers to mutually beneficial associations of fungi with roots. Mycorrhizas are very important in the uptake of nutrients such as P, N, K, Cu, Zn and Ca by plants especially in soil where these nutrients are present in low amount. P is the most limiting

nutrient in tropical soils, mycorrhizas are very important for improving P nutrition particularly for that soil. The thin mycorrhizal hyphae are able to penetrate soil pores (Kirkby and Mengel, 2009). Studies have shown that the heavily mycorrhizal root of cassava enables it to grow well in phosphate-deficient soils where other crops fail to grow (Wild, 1993). In alkaline soils, mycorrhiza have ability to prevent iron and manganese deficiencies.



Fig4. Showing mycorrhizal association

Source: Igual,2013

Shoot dry weight enhanced when the *Cassia* plants treated with mycorrhiza at a range of soil P levels than non-mycorrhizal treatment.

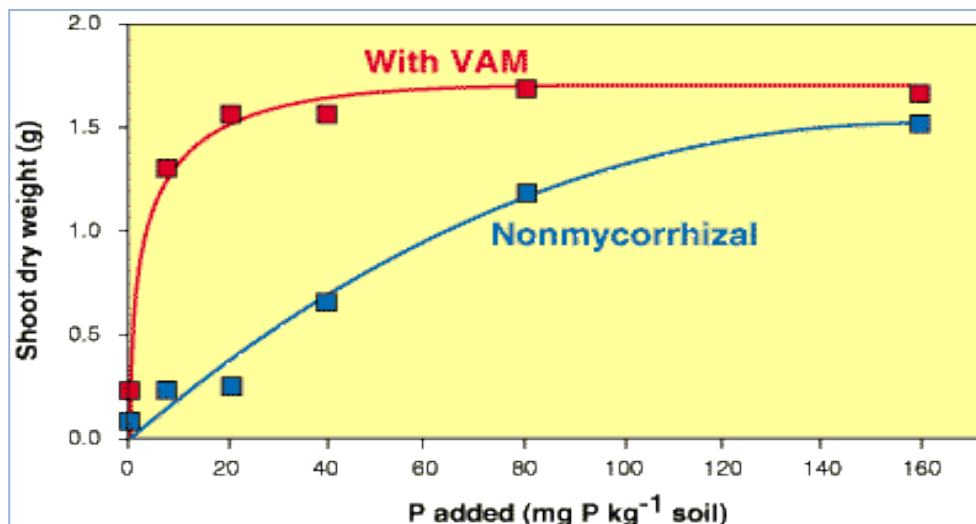


Fig5.Response curve was produced by growing mycorrhizal and nonmycorrhizal plants *Cassia* at a range of soil P levels.

Source: Sharma *et al.*, 2009

Table12. Mycorrhiza increase soil fertility by

Direct positive effects upon	Lessen negative effects from
<ul style="list-style-type: none"> • Uptake of immobile nutrients • Drought tolerance • Soil macroaggregates formation and stabilization • Soil organic matter 	<ul style="list-style-type: none"> • Root pathogens • Leaching loss of nutrients • Microbial immobilization of nutrients

Source: Sharma *et al.*, 2009

A study showed that wheat plant treated with Mycorrhiza absorbed higher rate of both macro and micro nutrients than non Mycorrhizal wheat plant (Table13).

Table13. AR ($\mu\text{g mg}^{-1}$) of macrolelements (P, Ca and Mg) and microelements (Cu, Zn and Mn) in mycorrhizal (+M) and nonmycorrhizal (-M) wheat plants

AM	Macro elements			Micro elements		
	P	Ca	Mg	Zn	Cu	Mn
-M	0.47	7.10	2.22	0.026	0.013	0.047
+M	0.58	7.20	2.24	0.044	0.015	0.049

Source: Cornejo *et al.*, 2008

Scientist showed that mycorrhiza are the effective nontoxifier of soil pollutants. There are several mechanisms such Phytoextraction and Phytostabilization are used by mycorrhiza during non toxification of pollutants such as As Cd, Cu, Pb etc. (Table14).

Table14. Mycorrhiza mediated nontoxification of some inorganic pollutants in soils their mechanisms.

Pollutant	Mechanism	Reference
As	Phytoextraction	Trotta <i>et al.</i> , 2006
Cd	Phytostabilization	owska <i>et al.</i> , 2012
Cu	Phytostabilization	Cheng <i>et al.</i> , 2007
Pb	Phytostabilization	Janouskova <i>et al.</i> , 2006

Source: British Microbiology Research Journal, 3(4): 724-742, 2013

Actinomycetes

Actinomycetes are necessary for the breakdown of certain components in organic matter. *Frankia* is a genus of actinomycetes. They form nitrogen-fixing root nodules with several woody plants of different families.

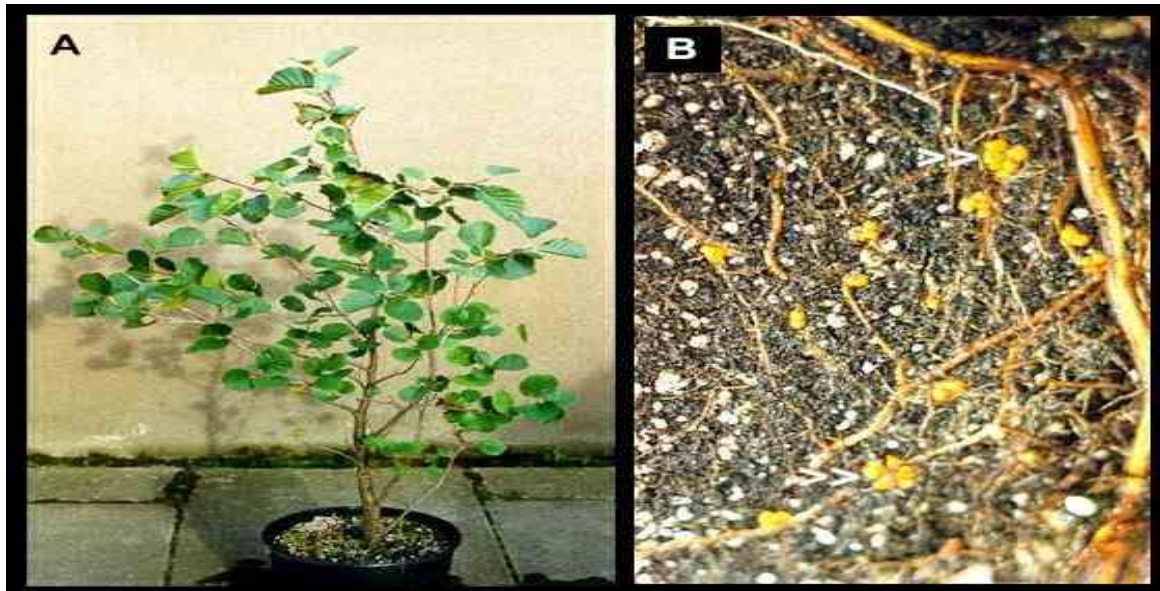


Fig6. Tree bearing the orange-yellow coloured nodules (arrowheads) containing *Frankia*

Source: Siddique., 2008

Microorganisms act as a pool of nutrients

Organic matter acts as nutrient reservoir for the plants. One of the most important pools of organic matter is the microbial biomass. The microbial biomass is a relatively available reservoir of plant nutrients such as nitrogen and phosphorus (Marumoto, 2006). Although the size of the microbial biomass is relatively small (e.g., its nitrogen content constitutes only 1 to 5 percent of the total organic nitrogen in soil), the nutrients within this pool are recycled rapidly within the soil profile, perhaps 8 to 10 times per year (Coleman, 2007). The amount of nitrogen in microbial biomass in agricultural soils ranges from 36 to 344 pounds per acre (40 to 385 kg/ha) (Paul and Vorone , 2006).

Microorganisms act as agents of change

The major groups of soil microorganisms are bacteria, fungi, and protozoa. Bacteria and fungi are responsible for the conversion of organic compounds to their mineral components, a process called mineralization. The mineralizers include microbes that split complex and large

plant molecules into smaller molecules. Many other agriculturally important functions are carried out by soil microorganisms. Important groups include the nitrifying bacteria (*Nitrosolobus*, *Nitrobacter*, *Nitrosomonas*) that are involved in the conversion of ammonium to nitrate. This conversion makes the nitrogen more available for plants. Mycorrhizal fungi make phosphorus more accessible to plants.

Involvement of Microorganisms in the Composting Process

Composting is a controlled decomposition of organic residues or waste to a state in which the composted material can be safely handled, stored and/or applied to land without adverse effects to the environment. It is a process that creates humus like organic materials. Composting mainly happens because of microbial activities. Microbes decompose organic materials through production of various extracellular enzymes such as peptidases, cellulases, hemicellulases and pectinases. (Table 15) presents results of the impacts of compost use on yields of different crops in Ethiopia.

Table 15. Average yields by treatment with compost in kg/ha for 5 crops

Crop	Average yields (kg/ha)	
	Check	Compost
Barley	1115	2349
Wheat	1228	2494
Maize	1760	3748
Sorghum	1338	2497
Field pea	1527	1964

Source: Kennedy., 2012

Role of microorganisms in removing heavy metals

Microorganisms play vital roles in removing heavy metals which are harmful for plant and also for human health. They detoxify heavy metals by breaking down their bonds and convert into simpler molecules which are nontoxic both for plants and animals. They also remove toxic heavy metals by direct uptake from the soil. They uptake heavy metals up to 10-170% of their dry weight (Table 16).

Table16. Microorganisms and uptake of heavy metals

<i>Microorganisms</i>	Elements	Uptake(% dry weight)
<i>Citrobacter spp.</i>	Co and Ni	25 and 13
<i>Bacillus spp.</i>	Cd	170
<i>Chlorella vulgaris</i>	Zn and Cu	15 and 14
<i>Rhizopusarrhizus</i>	Au	10
<i>Aspergillusniger</i>	Hg	58

Source: Kennedy *et al.*, 2013

Toxic pesticides degradation by microbes

A substantial decline of the residual lindane at different inoculum concentrations was observed within 0-2 weeks of incubation whereas the compound did not disappear from the uninoculated sterile control. Maximal pesticide depletion (56.0%) was observed at 4gcells Kg⁻¹soil (Table17)

Table17. Effect of inoculum size on the lindane removal by Streptomyces sp.in sterile soil samples, after 4 weeks of incubation

Inoculum size g Kg ⁻¹ soil	Lindane removal (%)
0.5	24.4
1.0	30.0
2.0	45.8
4.0	56.0

Source: Benson *et al.*, 2011

Biofertilizer and soil fertility improvement

Indiscriminate use of synthetic fertilizers has led to the pollution and contamination of the soil, has polluted water basins, destroyed microorganisms and friendly insects, making the crop more prone to diseases and reduced soil fertility. Depleting feedstock/fossil fuels (energy crisis) and increasing cost of fertilizers. This is becoming unaffordable by small and marginal farmers, depleting soil fertility due to widening gap between nutrient removal and supplies, growing concern about environmental hazards, increasing threat to sustainable agriculture. Besides above facts, the long term use of biofertilizers is economical, ecofriendly, more efficient, productive and accessible to marginal and small farmers over chemical fertilizers.

Table 18. Some commonly used biofertilizers with their active ingredient

Products	Active Ingredient	Use
ENRHIZO	<i>Rhizobium sp.</i>	Atmospheric nitrogen fixation for legumes.
ENFOSFO-P	<i>Bacillus polymyxa</i>	Converts non-available form of phosphorus to available form.
ENPOTASH	<i>Frateriiaaurantia</i>	Mobilize potash and make available to roots.
ENSULF	<i>Thiobacillusferrooxidans</i>	Enhances sulphur availability to plants.
ENZINC	<i>Thiobacillusthiooxidans</i>	Mobilize zinc and make available to roots.
ENFER	<i>Acidithiobacillus</i>	Mobilizes Ferrous in Soil

Source: Mohammadi, 2012

Seed yield increased drastically when rhizobia biofertilizer applied in the soybean field. Yield response increased as the P application increased.

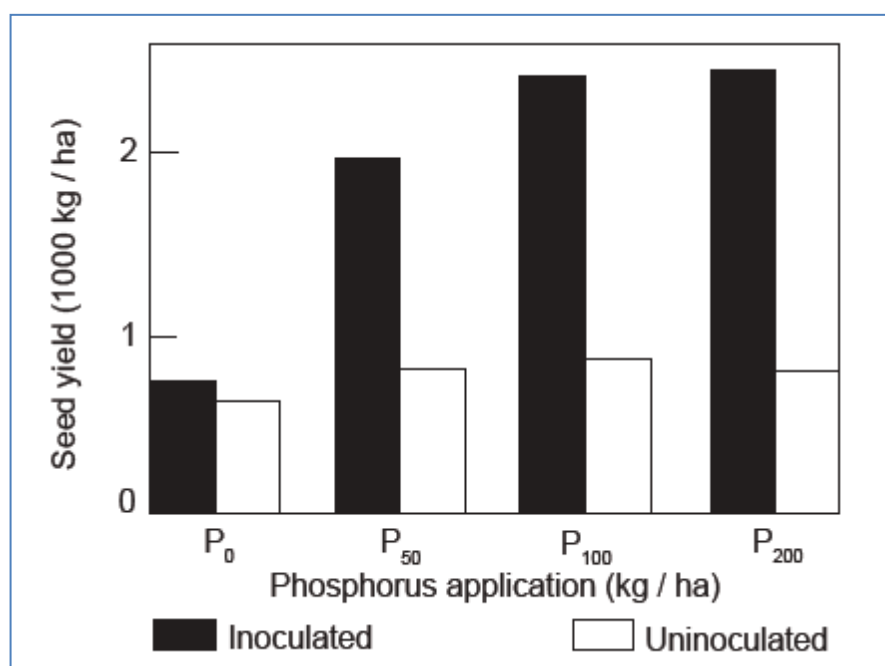


Fig7. Response of a soybean crop in Hawaii to inoculation with rhizobia biofertilizer with various levels of phosphorus fertilizer added.

Source :Uchida *et al.*,2008.

Biofertilizer Saves Chemical Fertilizer

A study conducted at Gujarat in India showed that a huge amount N and P saved when biofertilizer applied in the crop fields (Table19). 20-40 kg N/ha saved when *Azotobacterchroococcum* applied in sorghum field. The highest N (100 kg N) saved by the application of *Acetobacterdiazotrophicus* in sugarcane fields. 20-50kg P₂O₅ saved when *Bacilluscirculans* applied in the cowpea field (Table19).

Table19.Fertilizer saved when biofertilizer applied and their recommended crops

Type	Biofertilizer	Recommended Crop	Fertilizer Saving/ ha
N fixers	<i>Azollapinnata</i> (fresh)	Low land rice	30-50 kg N
	<i>Azotobacterchroococcum</i>	Pearl millet, sorghum	20-40 kg N
	BGA	Low land rice	30-50 kg N
	<i>Rhizobium spp.</i>	Pigeon pea and Chickpea	30-50 kg N
	<i>Azospirillumlipoferum</i>	Maize and sesame	20-40 kg N
	<i>Acetobacterdiazotrophicus</i>	Sugarcane	100 kg N
PSM	<i>Bacillus circulans</i>	Cow pea	20-50kg P ₂ O ₅
	<i>Bacillus brevis</i>	Wheat	
	<i>Bacillus coagulans</i>	Sorghum	
	<i>Torulopsoraglobosa</i>	Maize	

Source: Bastaet *al.*, 2010

Carbon Sequestration

Atmospheric carbon-di-oxide is increasing due to human sources of carbon dioxide emissions ,industrialrevolution,destroy of forest,burning of coal,gas,fossil fuel etc.So scientists are trying to invent the way of removal of extra carbon from the earth. And the carbon sequestration is the most accepted way. Plant use carbon dioxide from atmosphere for photosynthesis and emit carbon di oxide by respiration.

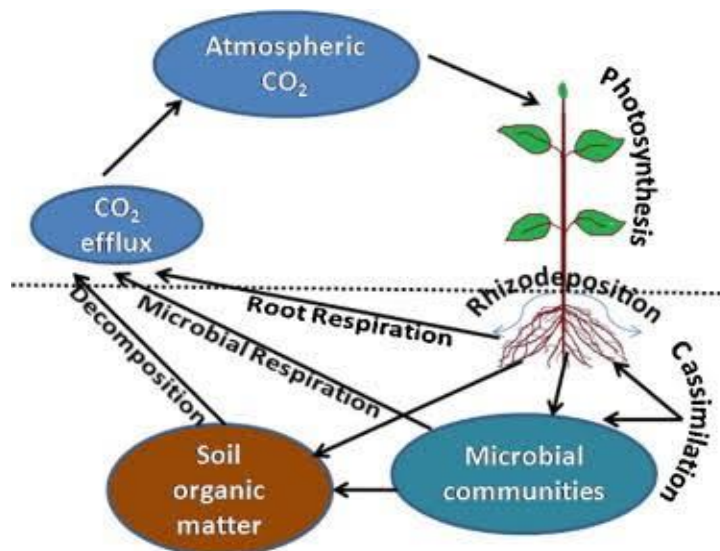


Fig8: Plant associated microorganism and carbon

When input of carbon in plant is higher than the output of carbon by plant then net carbon sequestration is occurred. In soil carbon is stored as soil organic carbon as a part of soil organic matter.

Soil carbon sequestration is a process in which carbon dioxide is removed from atmosphere and stored in the soil carbon pool. Carbon sequestration is mediated by soil microbes. Where the microbial mass is higher, carbon sequestration is higher in that soil than the soil which have less microbial mass. Microbial mass is influenced by land management, plant diversity and also influenced carbon storage in soil. Soil under organic farming condition accumulate more carbon than soil under conventional farming condition, even though organically farmed soil may be tilled frequently and incorporated with more less carbon rich residues.

It also found that higher biodiversity of plants increase the root mass which is responsible for higher soil microbes. And higher soil microbes accumulate higher carbon than bare land or less vegetated area. (source; USDA-funded research at Michigan State University's Kellogg Biological Station)

Carbon Sequester Micorrhizal fungi

These fungi are associated with plant roots and get sugar from root exudation. They use sugars and help plants to uptake more water and nutrients. By eating sugars, micorrhizal fungi deposits carbon containing residues in surrounding soil. Ectomycorrhizal fungi help to store 50% - 70% carbon in soil (Averill, c, et al)



Fig9: Fruting body ectomycorrhizal fungi from genus *Amantia*

Sequester Bacteria There are some aerobic bacteria such as Knallgasbacteria (chemolithoautotrophs). As a carbon source some anaerobic bacteria and archaea (e.g., acetogenic bacteria, carbon monoxide utilizer, methanogenogens) use carbon dioxide and can fix C.

Humification

If photosynthesis is higher than the plant and microbes respiration then more carbon will get stored in humus by humification process. Humus is a stable product which is produced by organic matter degradation by microbes. Carbon can persist in carbon rich humus for long periods.

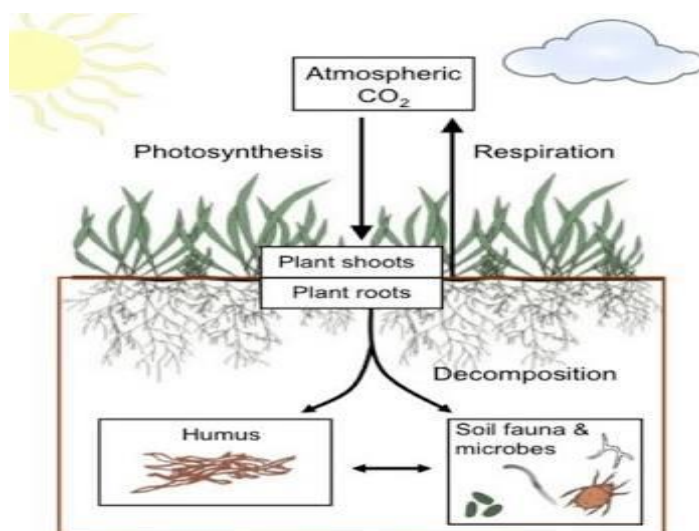


Fig10: Soil carbon storage by Humification

Challenges Faced in Exploiting Soil Microorganisms

There are a number of factors that derail maximization of beneficial microbial activities. Apart from climatic factors, there are a number of anthropogenic or social factors that affect biological functions and soil productivity. Chianu *et al.* noted the following some of the main challenges faced in exploiting beneficial activities of soil microorganisms for improving soil fertility and productivity:

- Absence or very weak institutions, policy and budgetary support for research.
- Limited knowledge of inoculation.
- Poorly developed marketing channels and infrastructure and limited involvement of the private sector in the distribution of inoculants.
- Limited farmer awareness about and access to inoculants.
- Lack of development of technology in this field.

CHAPTER IV

CONCLUSIONS

From this review it is noted that soil microorganisms have a huge contribution in soil fertility and carbon sequestration. This is achieved through a number of ways. Biological nitrogen fixation is an economically attractive and ecologically sound route for augmenting nutrient supply. The commonly reported *Rhizobium*/legume symbiosis contributes substantial amounts of biologically fixed nitrogen to cropping systems and significantly benefits crops that follow in rotation. Soil microorganisms such as bacteria and fungi contribute to plant phosphorus nutrition through solubilization of fixed or precipitated phosphorus from complexes with Al and Fe in acidic soils and calcium complexes in alkaline soils. Phosphate solubilizing rhizospheric bacteria has a high potential to be used in the management of P deficient soils.

Mycorrhizal associations are reported to contribute to plant phosphorus nutrition through increasing root surface area for soil exploration, production of phosphorus solubilizing enzymes and organic acids. Mycorrhizal fungi and bacteria also solubilize other nutrients such as zinc, copper, and calcium. Microorganisms also contribute to soil fertility improvements through their roles in composting. Soil microorganisms are currently isolated, studied and packaged as biofertilizers and used to supplement chemical fertilizers. The relevance of biofertilizers is increasing rapidly since chemical fertilizers (nitrogenous fertilizers) damage the environment. In contrast, biofertilizers lead to soil enrichment and are compatible with long-term sustainability. Further they are ecofriendly and pose no danger to the environment. Again, SOC is a vital component of soil and has important effects on terrestrial ecosystem. From interaction among the dynamic ecological processes of photosynthesis, respiration, decomposition which are mediated by microbes, net carbon is stored in soil. Carbon dioxide is increased greatly by human activities, but now human activities provide an opportunity for sequestering carbon back into soil. Carbon sequestration also improves soil quality by maintaining and increasing soil organic matter, adds to soil fertility, water retention and crop production. It can be noted that thorough exploitation of microbial activities can contribute to balanced fertilization and carbon storage for long term which help in improving long term soil health.

REFERENCES

- Abd-Alla, M. H. (1994). Phosphatases and the utilization of organic phosphorus by *Rhizobium leguminosarum* biovar *viceae*. *Letters in Applied Microbiology*, 18(5), 294-296.
- Afzal, A., & Bano, A. (2008). *Rhizobium* and phosphate solubilizing bacteria improve the yield and phosphorus uptake in wheat (*Triticum aestivum*). *Int J Agric Biol*, 10(1), 85-88.
- Alam S., S. Khalil, N. Ayub and M. Rashid. 2012. *In vitro* solubilization of inorganic phosphate by phosphate solubilizing microorganism (PSM) from maize rhizosphere. *Intl. J. Agric. Biol.* 4:454-458.
- Basta N.T., Ryan J. A. and Chaney R. L. (2010). Trace element chemistry in residual-treated soil: key concepts and metal bioavailability. *Journal of Environmental Quality* 34(1):49–63.
- Benson DR, Brooks JM, Huang Y, Bickhart DM, Mastroruozio JE. 2011. The biology of *frankia* sp. Strains in the post-Genome Era. *mol. plant –microbe interact.* Vol(24):1310-1316.
- Biesboer D, Binford MW, Kolata A. 2009. Nitrogen Fixation in Soils and Canals of Rehabilitated Raised-Fields of the Bolivian Altiplan. *Biotropica*, Vol. 31, No.(2), pp.
- Bocchi S, Malgioglio A. 2010. *Azolla-Anabaena* as a Biofertilizer for Rice Paddy Fields in the Po Valley, a Temperate Rice Area in Northern Italy. *International Journal of Agronomy*. vol(2010):1-5.
- Bohloul BB, Ladaha JK, Garrity DP, Gedrgé T. 2012. Biological nitrogen fixation for sustainable agriculture; A perspective. *Plant and soil*. V;141:1-11.
- Cottenie, A., 2010. Soil and plant testing as a basis of fertilizer recommendations. *FAO Soils Bulletin* 38/2, pp. 94-100.
- Damir O, Mladen P, Božidar S, Srđan N. 2011. Cultivation of the bacterium *Azotobacter chroococcum* for preparation of biofertilizers. *African Journal of Biotechnology* Vol. 10(16), pp: 3104 -3111.
- Fallah, A Schillinger N. 2009. Abundance and distribution of phosphate solubilizing bacteria and fungi in some soil samples from north of Iran. 18th World Congress of Soil Science, July 9-15, 2006, Philadelphia, Pennsylvania, USA.

- Fankem, H., D. Nwaga, A. Deubel, L. Dieng, W. Merbach and F. X. Etoa. 2006. Occurrence and functioning of phosphate solubilizing microorganisms from oil palm tree (*Elaeisguineensis*) rhizosphere in Cameroon. *African J. Biotech.* 5:2450-2460.
- Hilda, R. and Hayat. 2007. Phosphate solubilizing bacteria and their role in plant growth promotion. *Biotech. Adv.* 17:319-359.
- Hinsinger, P. and Sharma N. 2009. Bioavailability of soil inorganic P in the rhizosphere as affected by rootinduced chemical changes: a review. *Plant Soil* 237:173-195.
- Igual, J. M., A. Valverde, E. Cervantes and E. Velázquez. 2011. Phosphate-solubilizing bacteria as inoculants for agriculture: use of updated molecular techniques in their study. *Agronomie*.21:561-568.
- Kennedy AC, Khan TL, Schillinger WF. 2012. Soil and crop management effects on soil microbiology. In: Magdoff F, Weil RR, editors. *Soil Organic Matter in Sustainable Agriculture*. CRC Press.;295-326.
- Matthew, C.J., M.K. Bjorkman, M.K. David, A.M. Saito and P.J. Zehr, 2008.Regional distributions of nitrogen-fixing bacteria in the Pacific Ocean.*Limnol.Oceanogr*, 53: 63-77.
- Mohod, S.; Gupta, D. N.Akhtar, and Chavan, A. S. 2006.Effects of P Solubilizing organisms on yield and N uptake by rice. In: *Journal of Maharashtra Agricultural Universities*. Vol. 16, no. 2; p. 229-231.
- Mortvedt, J. J. , Giordano, P. M.,Cornejo Lindsay, W. L.,2008. Micronutrients in agriculture. *Soil Sci. Soc. Am. Inc. Madison, Wisconsin, USA*.
- NilabjaGhosh N. 2012. Promoting Bio-fertilizers in Indian Agriculture.nstitute of Economic Growth for the Ministry of Agriculture, Government of India.
- Omar, S. A.2009. The role of rock-phosphate-solubilizing fungi and vesicular–arbuscular mycorrhiza (VAM) in growth of wheat plants fertilized with rock phosphate. *World J. Microbiol.Biotechnol.* 14:211-218
- Saikia SP, Jain V and Qureshi. 2007. Biological nitrogen fixation with non legumes: An achievable target or a dogma? *Current Science*, Vol. 92, No.3:317-322.
- Santi C, Bogusz D, Franche C. 2013. Biological nitrogen fixation in non-legume plants. *Annals of Botany* .Vol.111: 743 –767
- Simon, T., 2013.Utilization of biological nitrogen fixation for soil evaluation. *Plant Soil Environ.*, 49: 359-363.

- Subbarao, N. S. 2007. Phosphate solubilizing micro-organism. In: Biofertilizer in agriculture forestry. Regional Biofert. Dev. Centre, Hissar, India. pp. 133-142.
- De Graaff, M. A., Classen, A. T., Castro, H. F., & Schadt, C. W. (2010). Labile soil carbon inputs mediate the soil microbial community composition and plant residue decomposition rates. *New Phytologist*, 188(4), 1055-1064
- De Graaff, M. A., Classen, A. T., Castro, H. F., & Schadt, C. W. (2010). Labile soil carbon inputs mediate the soil microbial community composition and plant residue decomposition rates. *New Phytologist*, 188(4), 1055-1064.
- Kalbitz, K., & Kaiser, K. (2008). Contribution of dissolved organic matter to carbon storage in forest mineral soils. *Journal of Plant Nutrition and Soil Science*, 171(1), 52-60.
- Clemmensen, K. E., Bahr, A., Ovaskainen, O., Dahlberg, A., Ekblad, A., Wallander, H., ...& Lindahl, B. D. (2013). Roots and associated fungi drive long-term carbon sequestration in boreal forest. *Science*, 339(6127), 1615-1618.